

# Instructions for Use

## Reticulum Control Slides

CATALOG NUMBER	DESCRIPTION	UNIT OF MEASUREMENT
CS024-PS-25	Reticulum Control Slides, Adhesion, 1 stained/24 unstained	1 set
CS024-PS-100	Reticulum Control Slides, Adhesion, 1 stained/99 unstained	1 set

### INTENDED USE

Qualitative control for reticulum.

### TISSUE

Liver

### FIXATION

10% Buffered Formalin

### TECHNICAL INFORMATION

Paraffin sections cut at 5 microns

### USE WITH POLYSCIENTIFIC GORDON SWEET'S RETICULUM STAIN KIT

Kit Size	Part Number
8 oz	K082

### POLYSCIENTIFIC GORDON SWEET'S RETICULUM STAIN PROCEDURE

1. Deparaffinize and hydrate to running DI water.
2. Oxidize in Acidified Potassium Permanganate Working Solution for 5 minutes.
3. Wash well in water for 3 minutes.
4. Bleach in Oxalic Acid 1% Aqueous until there is no trace of brown color.
5. Rinse well in tap water for 5 minutes.
6. Place in Ferric Ammonium Sulfate 2.5% Aqueous for 15 minutes. No crystals should be at the bottom of staining dish. If so, fresh solution must be made.
7. Rinse in several changes of DI water. Leave slides in DI water until staining jars have been prepared in the following order:
  - a. Silver Nitrate Working Solution, 1 jar
  - b. Distilled Water, 3 jars
  - c. Formaldehyde 37%, 1 jar
  - d. Distilled Water, 3 jars
8. Place in Silver Nitrate Working Solution for 10 dips of more.
9. Rinse well in DI water for 3 changes, 10 dips each. Water must be changed after each slide.
10. Place in Formaldehyde, 37% for 3 dips. The reaction takes place immediately.  
DO NOT LEAVE SLIDES IN THE SOLUTION
11. Rinse will in DI water for 3 changes, 10 dips each. Check under microscope. Fibers should be dark black and background colorless to yellowish-brown. If reticulum fibers are not black, slide should be rinsed for at least 3 minutes in running water and repeated from Step 8.

12. Tone in Gold Chloride 0.5% Aqueous for 1-12 dips. Do not overdo or slide will turn a reddish color.
13. Rinse well in water for 5 minutes.
14. Counterstain if desired in Neutral Red 1% Aqueous.
15. Rinse in water for 15 dips.
16. Dehydrate in 95% Alcohol and Absolute Alcohol, 2 changes each.
17. Clear in Xylene, 2 changes each.
18. Coverslip with Acrymount Plus or any other acceptable mounting medium.

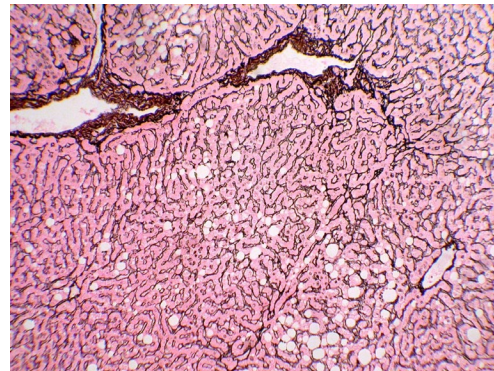
### RESULTS

Reticulum Fibers – Gray to black

Nuclei – Pink to red

This protocol is a recommendation only. Modifications may be necessary.

Please contact [productsupport@statlab.com](mailto:productsupport@statlab.com) with any additional questions.



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